



# Biological control efforts for the management of Fall Armyworm in India

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**ICAR- National Bureau of Agricultural Insect Resources, Bengaluru, INDIA**

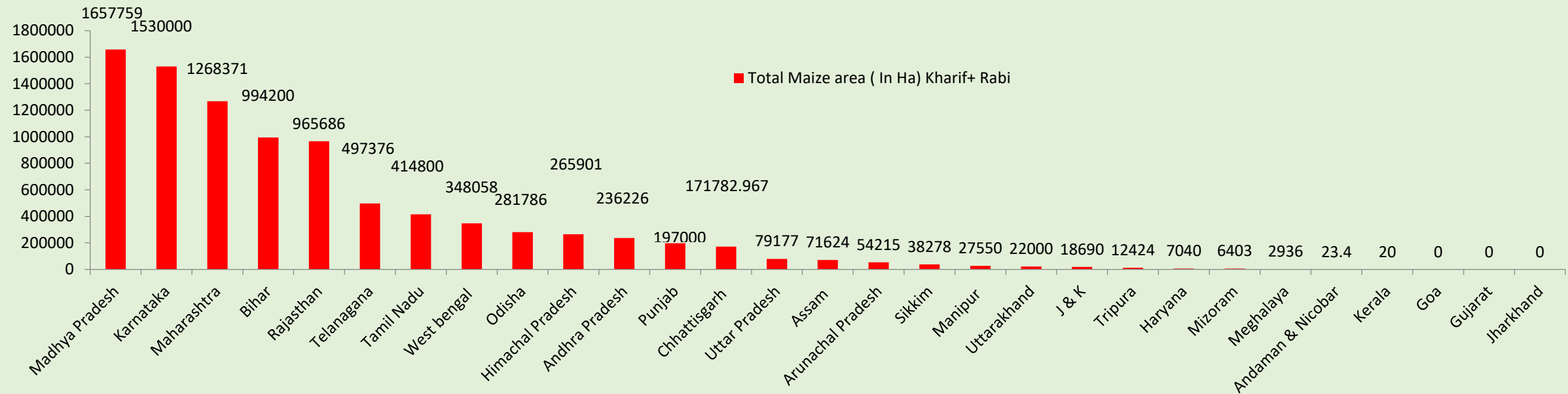
## the facts....

- FAW was first recorded in **May 2018** in India in southern most part (Shylesha et al. 2018) and later spread to other states (except Kashmir).
- First recorded on maize, later on sorghum
- Occasionally on sugarcane, cotton, rice, ginger etc.
- Maize cobs also damaged occasionally.
- **Incidence in 2018:** upto 70% plant damage.
- **Incidence in 2022-23:** low to moderate (6-20%) in all states as per AICRP-BC report.



# Maize Cultivation Scenario in India (2022-23)

Total Maize area ( In Ha) Kharif+ Rabi



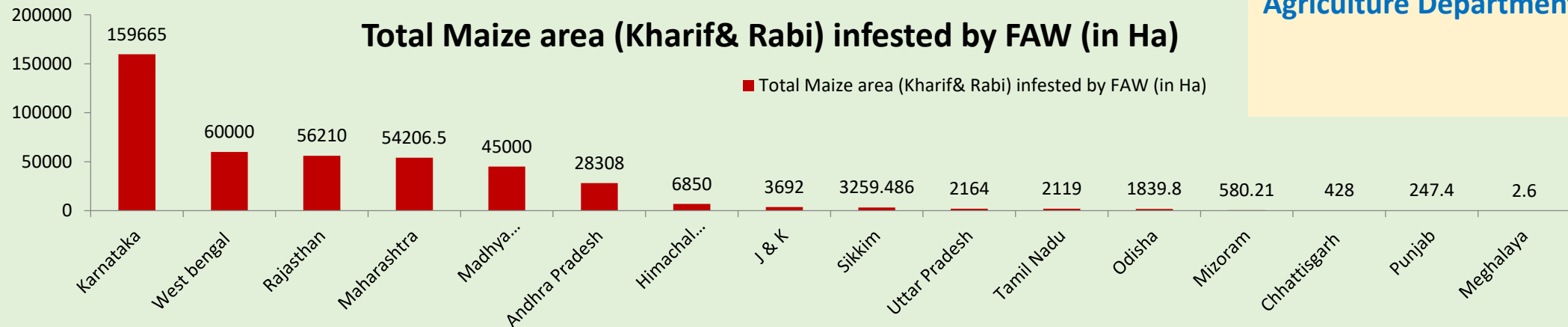
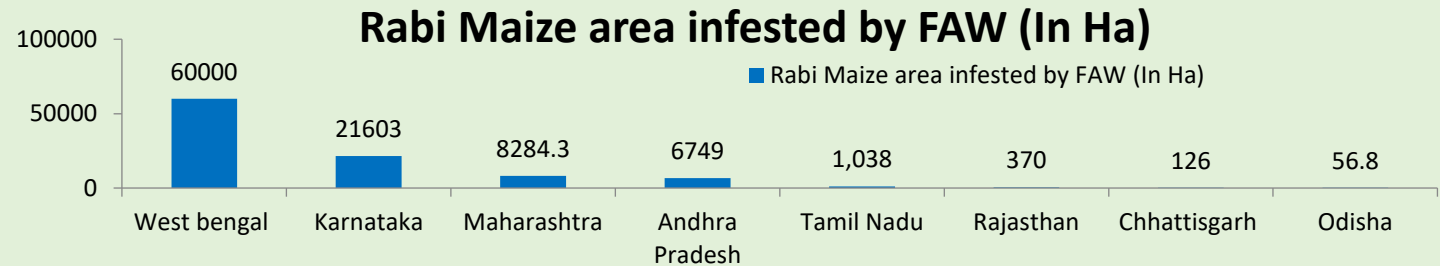
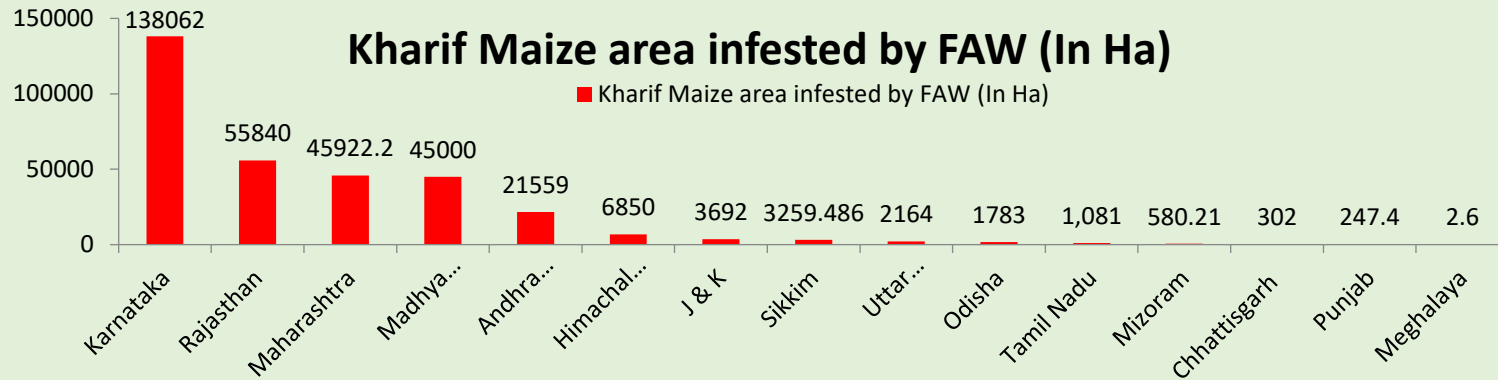
**Kharif (Sowing in May-June) - 6.80 M Ha (74%)**

**Rabi - ( Sowing in Oct-Nov) 2.36 M Ha (26%)**

**Total area - 9.17 M Ha**

(Source: Data Submitted by State Agriculture Departments)

# Maize Area (Ha) Infested by FAW in India (2022-23)



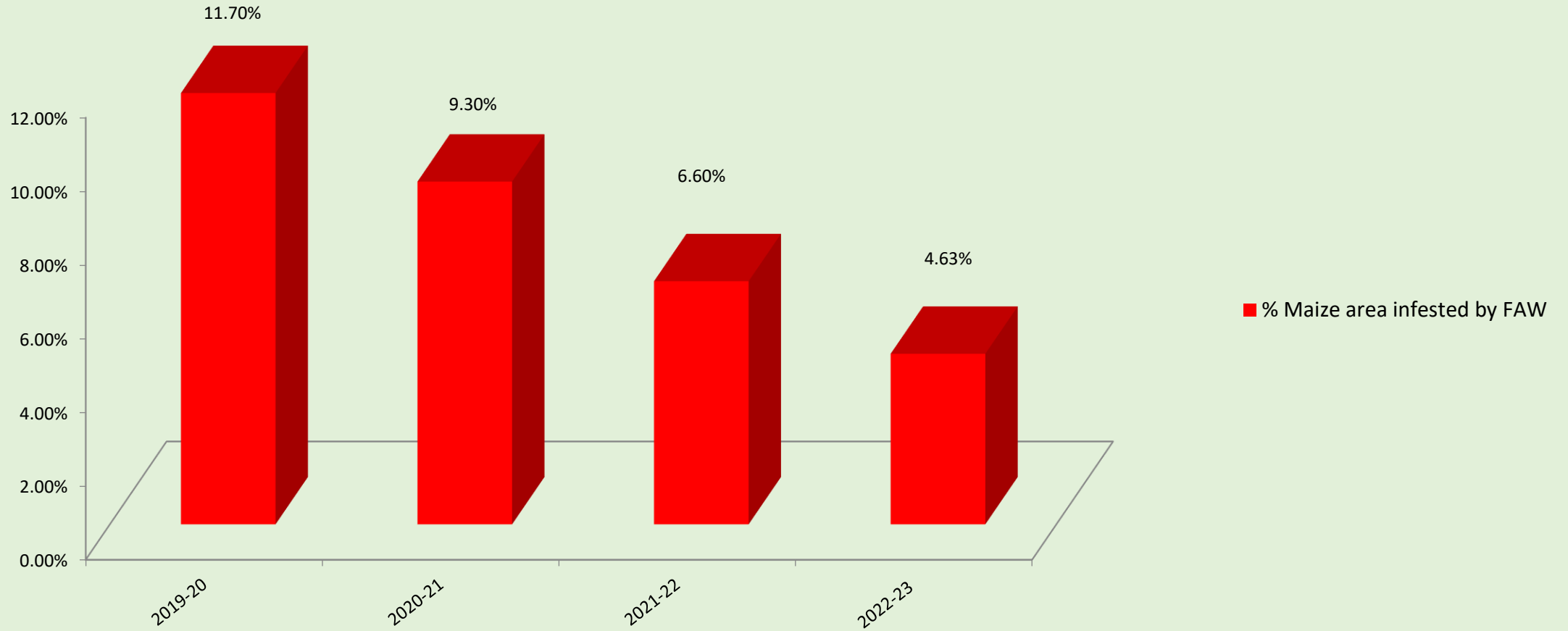
**Kharif (Sowing in May-June) –  
0.326 Million ha ( 4.79% area of  
Kharif)**

**Rabi - ( Sowing in oct-Nov)  
0.098 Million ha (4.15 % area of  
Rabi)**

**Total 0.424 Million ha ( 4.63% of  
Total Maize area)**

**Source: Data Submitted by State  
Agriculture Departments**

# Percent maize Area infested by FAW in India (2019--23)












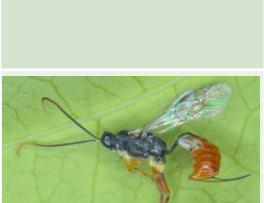



# Surveys conducted in states across India










# Natural enemies recorded on fall armyworm (FAW) in India

	Scientific name	Taxonomic placement	Biological attribute	Collection locality (States)	Reference
	<i>Telenomus remus</i>	Hymenoptera: Platygastridae	Egg parasitoid	Karnataka and Tamil Nadu	Shylesha et al. 2018
	<i>Trichogramma chilonis</i>	Hymenoptera: Trichogrammatidae	Egg parasitoid	Karnataka	Shylesha et al. 2018
	<i>Chelonus formosanus</i> Sonan	Hymenoptera: Braconidae	Egg-larval parasitoid	Karnataka and Telangana	Gupta et al. 2020
	<i>Coccygidium transcaspicum</i>	Hymenoptera: Braconidae	Larval parasitoid	Karnataka and Telangana	Gupta et al. 2020
	<i>Cotesia ruficrus</i> (Haliday)	Hymenoptera: Braconidae	Larval parasitoid	Karnataka, Tamil Nadu, Rajasthan, Uttar Pradesh, Punjab, Meghalaya	Gupta et al. 2019

	<b><i>Glyptapanteles creatonoti</i> (Viereck)</b>	Hymenoptera: Braconidae	Larval parasitoid	Karnataka	Shylesha et al. 2018
	<b><i>Campoletis chlorideae</i> Uchida</b>	Hymenoptera: Ichneumonidae	Larval parasitoid	Karnataka	Shylesha et al. 2018 and Sharanabasappa et al. 2019
	<b><i>Eriborus</i> sp.</b>	Hymenoptera: Ichneumonidae	Larval parasitoid	Karnataka	Sharanabasappa et al. 2019
	<b><i>Odontepyrus</i> sp.</b>	Hymenoptera: Bethylidae	Larval parasitoid	Tamil Nadu	Sharanabasappa et al. 2019
	<b><i>Phanerotoma</i> sp.</b>	Hymenoptera: Braconidae	Larval parasitoid	Karnataka	Ballal et al. 2021
	<b><i>Exorista sorbillans</i> (Wiedemann)</b>	Diptera: Tachinidae	Larval parasitoid	Karnataka	Sharanabasappa et al. 2019



	<b><i>Forficula</i> sp.</b>	<b>Dermaptera: Forficulidae</b>	<b>Predator</b>	<b>Karnataka</b>	<b>Shylesha et al. 2018 and Sharanabasappa et al. 2019</b>
	<b><i>Harmonia octomaculata</i> (Fabricius):</b>	<b>Coleoptera: Coccinellidae</b>	<b>Predator</b>	<b>Karnataka</b>	<b>Sharanabasappa et al. 2019</b>
	<b><i>Coccinella transversalis</i> Fabricius</b>	<b>Coleoptera: Coccinellidae</b>	<b>Predator</b>	<b>Karnataka</b>	<b>Sharanabasappa et al. 2019</b>
	<b><i>Andrallus spinidens</i> (Fabricius)</b>	<b>Hemiptera: Pentatomidae</b>	<b>Predator</b>	<b>Karnataka</b>	<b>Firake &amp; Behere 2020</b>
	<b><i>Eocanthecona furcellata</i></b>	<b>Hemiptera: Pentatomidae</b>	<b>Predator</b>	<b>Karnataka</b>	<b>Keerthi et al. 2020</b>

# Egg and larval parasitoids

**First year of  
invasion 2018**

1. *Trichogramma chilonis* Ishii
2. *Telenomus* nr. *remus*
3. *Chelonus formosanus* Sonan
4. *Campoletis chloridae* Uchida
5. *Cotesia ruficrus* (Haliday)
6. *Glyptapanteles creatonoti* (Viereck)



# Gradually expanding native parasitoid bank..... post 2018

## Two-Three years of invasion

1. *Coccygidium transcaspicum* (Kok.), 2. *Phanerotoma* sp., 3. *Aleiodes* sp., 4. *Meteorus pulchricornis* (Wes.), 5. *Microplitis manilae* (Ashmead), 6. *Microplitis demolitor* (Wilk.), 7. *Netelia* sp., 8. *Metopius rufus* (Ashmead), 9. *Temelucha* sp., 10. *Ichneumon promissorius*.



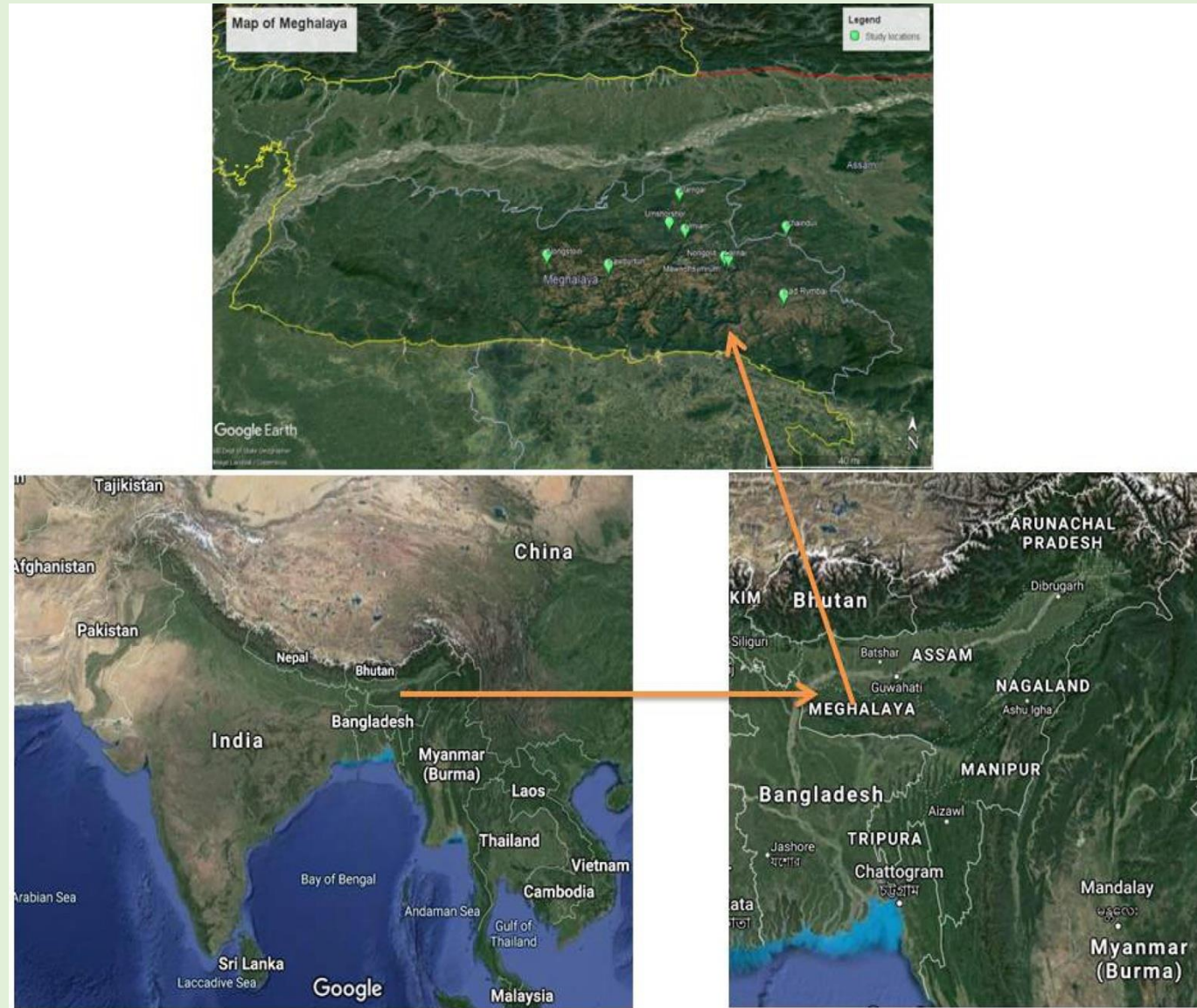


# Natural percent parasitism.....

Parasitoids	Percent parasitization	
	2018-19	2019-20
<i>Trichogramma chilonis</i>	3.7 - 12.43	15.81 - 23.87
<i>Telenomus remus</i>	1.8 - 9.4	8.0 - 11.6
<i>Chelonus</i> spp.	0.2 - 1.4	4.91 - 21.38
<i>Cotesia</i> sp.	-	0.14
<i>Campoletis chlorideae</i>	-	0.29
<i>Exorista xanthaspis</i>	-	1.86 - 6.63



## Experiences from NE India.....



**In Meghalaya (N-E India) : 56.6% to 73.1% of larvae were found to be parasitized /infected with entomopathogens.**

**34 different locations representing five major districts (viz., East Jaintia Hills, West Khasi Hills, East Khasi Hills, West Jaintia Hills and Ri Bhoi)**

**Firake and Behere, 2020**



# Genetic diversity of invasive FAW populations in India

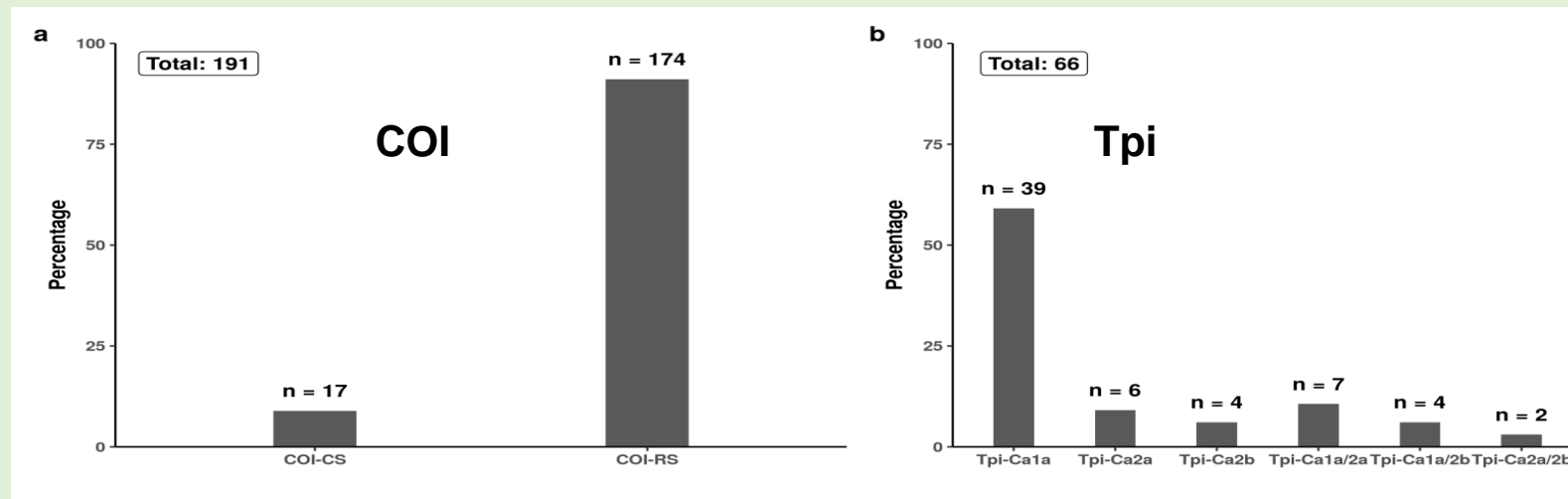
- Two morphologically identical strains, designated as majority strain (*Tpi*-Corn & *COI*-Rice) and minority strain (*Tpi*-Corn & *COI*-Corn) most consistently found on Maize and less frequently in Sorghum, Finger Millet, Pearl Millet etc.
- Genetic diversity of the two strains was studied based on *Cytochrome c-oxidase subunit-1* (mitochondria) & *Triosephosphate isomerase* (nuclear) markers.
- The same trend was found in other invaded countries.





# Genetic diversity of invasive FAW populations in India

- Analyzed 191 mitochondrial COI gene sequences of FAW specimens from 101 different regions of India & Nepal
- In addition, 66 nuclear *Tpi* gene sequences were also analyzed which is considered as a useful marker to ascertain the association of host plants in FAW.



Distribution of FAW haplotypes in India (*COI* & *Tpi*)

## scientific reports

OPEN

### Population structure and genetic diversity of invasive Fall Armyworm after 2 years of introduction in India

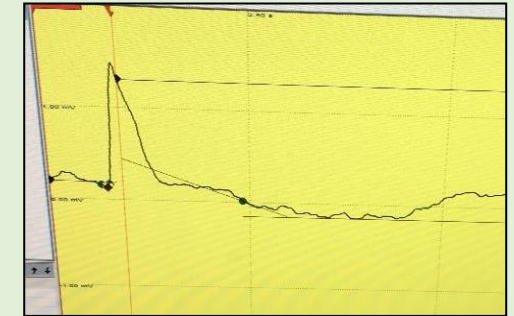
N. Nayyar<sup>1</sup>, R. G. Gracy<sup>2</sup>, T. R. Ashika<sup>1,3</sup>, G. Mohan<sup>1,3</sup>, R. S. Swathi<sup>1</sup>, M. Mohan<sup>1</sup>, M. Chaudhary<sup>2</sup>, N. Bakthavatsalam<sup>1</sup> & T. Venkatesan<sup>1,2</sup>

Fall Armyworm (FAW), *Spodoptera frugiperda*, is a polyphagous pest capable of feeding over 80 plant species and was indigenous to the Western Hemisphere. Within a span of 4 years, FAW has established itself throughout most of the regions in Africa and Asia causing significant losses in maize production. Owing to its revamped distribution range, it would be prudent to analyze the ensuing genetic changes and study the emerging phylogeographic patterns across the world. In this regard, we would like to

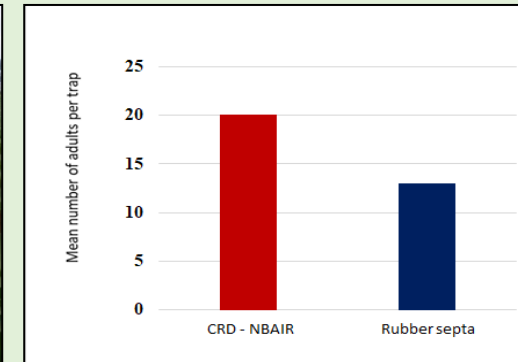
- 100% of the population exhibited 'Corn Strain' haplotype on the basis of *Tpi* gene sequences.
- Thus, *Tpi* gene marker was more accurate in prediction of host association in India. Similar trend was observed in FAW from Africa & Asia, suggesting a common origin for invasive populations in all these places.

# Controlled release matrix for delivery of FAW, *Spodoptera frugiperda* pheromone

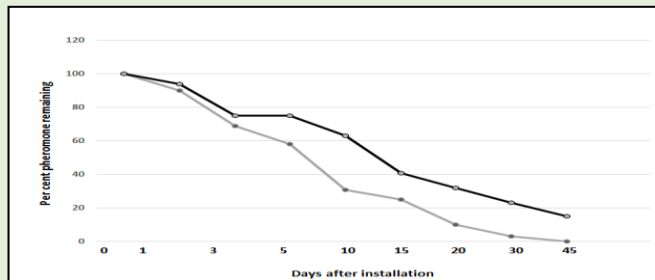
- The Antennal response of adult *S. frugiperda* males to (Z)- 9-Tetradecen-1-ol acetate (as major component) was established by electroantennography. This confirms the physiological response of the antennal neurons to pheromone.
- Field trials using the **NBAIR controlled release dispenser** at three locations trapped over  $20 \pm 3.13$  adult males per trap as compared rubber septa that trapped less than  $13 \pm 2.82$  moths per trap.
- Controlled released dispenser is effective for over 45 days.
- Identity of *S. frugiperda* adults trapped were confirmed by molecular and morphological techniques
- The technology developed has been commercialized to three firms



EAG trace of FAW adult



Field efficacy of FAW pheromone



Line in black : Pheromone loaded in controlled release matrix  
Line in grey : Pheromone loaded in rubber septa



Trapping FAW adult males



*S. frugiperda* genitalia dissected to confirm the sp. trapped



# Microbial pathogens on FAW in India



## Quantification of Natural occurrence of microbial pathogens of FAW in Karnataka (India)

Fungal pathogens : 74.07 %

Bacterial pathogens : 9.87%

NPV : 13.58%

Others : 3.70%

- Natural epizootics of *Metarhizium rileyi* (= *Nomuraea rileyi*) were reported to cause significant larval mortality ranging from 1.87% to 18.30% in Karnataka.
- Natural epizootics of an NPV of *S. frugiperda* have been reported from Gujarat in Western India (Raghunandan et al., 2019).



# *Metarhizium anisopliae* ICAR-NBAIR Ma 35 for the management FAW

- Talc/oil based formulation was developed
- Field trials- 3 Foliar sprays @ 20, 30 and 40 days after sowing (10 days interval)
- **Locations:** Andhra Pradesh , Karnataka, Gujarat, Tamil Nadu
- Field trials - **50-76% reduction** in the plant damage caused by the FAW
- Yield enhancement- **20-40%**



FAW mortality under lab conditions



Ma35 spraying in maize plots



Ma-35 dead cadaver after foliar application

# Consortia of bacteria for the Management of FAW in hybrid Maize under Field Conditions

- A consortium of *Pseudomonas fluorescens* (NBAIR-PFDWD) and *Bacillus albus* (NBAIR-BATP) was found to be effective against FAW.
- More than **80 %** reduction in FAW infestation in maize.
- **One hundred per cent recovery** observed on maize plants infested with fall armyworm after four sprays of bacterial consortium.
- Plant growth promotion activity also observed.
- Yield enhancement: **30-45%**



Bacterial biopesticides sprayed maize field



Maize field inspection by experts

# Aqueous formulation of *Spodoptera frugiperda* nucleopolyhedrovirus (SpfrNPV ICAR -NBAIR1) for the management of FAW



**Shatpada FAW Kill**

- SpfrNPV is a host specific NPV and is effective against FAW
- Shelf life: 2 years
- Occlusion Bodies count in the formulation :  $1 \times 10^9$  OBs/ml
- Technology transferred for wide area demonstration.





# *Spodoptera frugiperda* NPV ICAR-NBAIR1 (FAW Kill)



Size of the Occlusion body	1.64 $\mu\text{m}$
Shape of the Occlusion body	Tetrahedral
Molecular confirmation	NCBI GenBank acc.no. MT422725 (Polyhedrin gene)
Laboratory larval mortality	80%
LC <sub>50</sub> Value	4.12 OB/mm <sup>2</sup>
Pest reduction recorded in field evaluation	72%

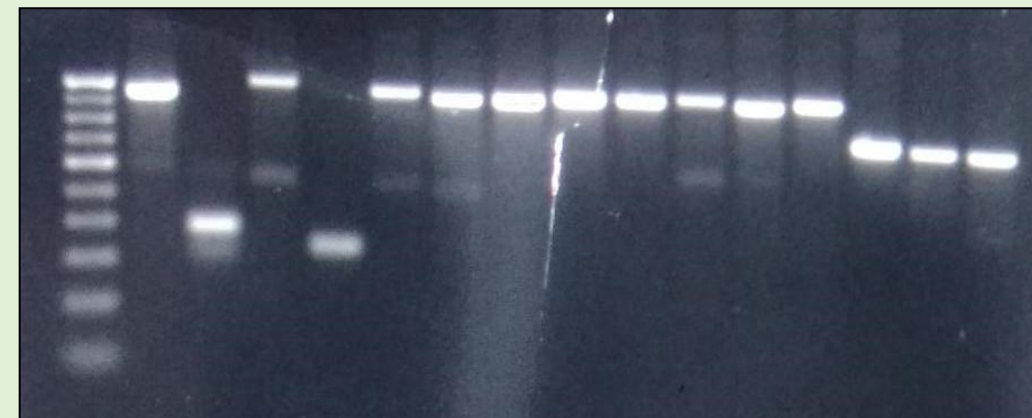
# Shatpada Armour (NBAIR Bt25)

## Bioefficacy of *B. thuringiensis* strain NBAIR BT25 against *Spodoptera frugiperda* after four days of inoculation

Isolates	LC50 (µg/ml)	95%FL	LC90 (µg/ml)	95%FL	Slope± SE	Chi-Square value	P-value
Bt25	42.92	33.30- 54.63	132.49	96.26-221.9	2.61± 0.38	4.29	0.23

## Field efficacy of *B. thuringiensis* strain NBAIR BT25 at multi-locations

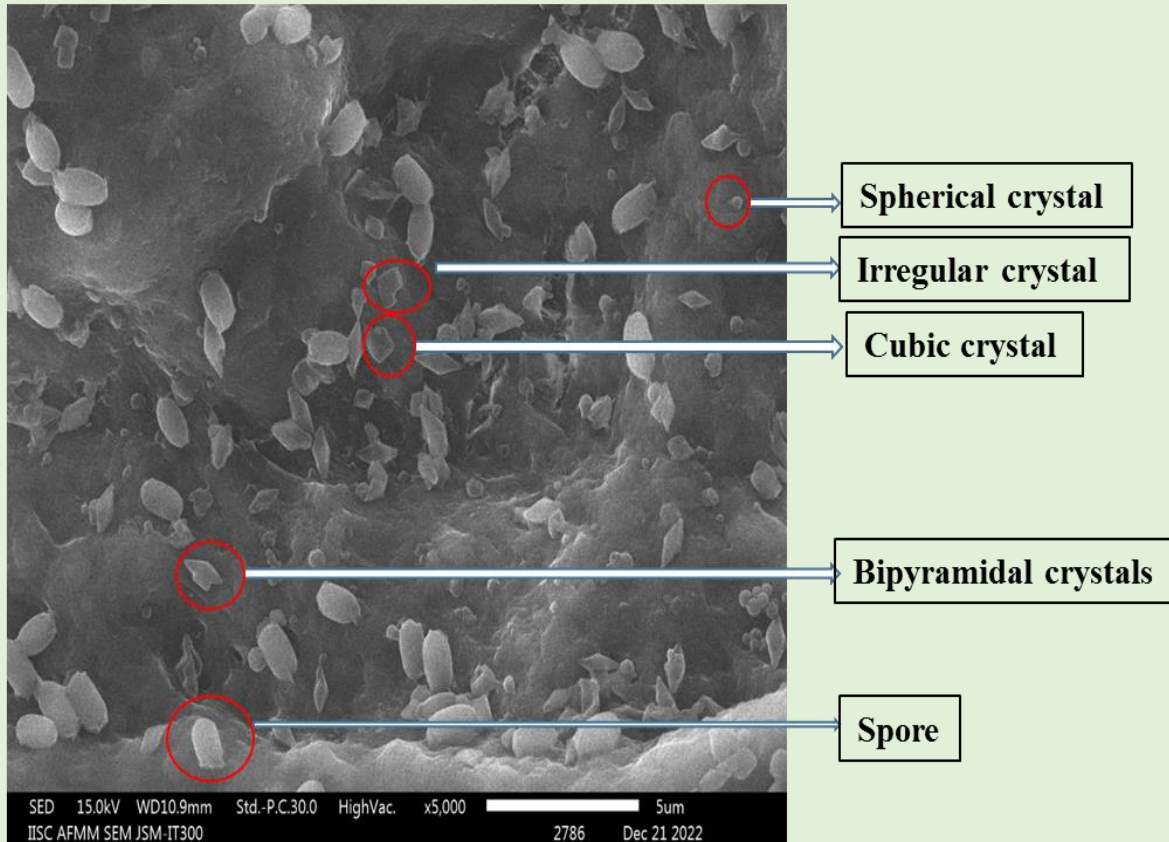
Multi-environment phenotyping of *B. thuringiensis* strain against FAW of maize was done in the year 2019-2020 where mean of average reduction of FAW larvae by NBAIR BT25 for different locations in 2019 was 57.47 % and in 2020 was 58.58 %.



Gel picture showing the insecticidal genes profiling of NBAIR BT25. Lane M: Ladder (100bp). Lane 1: *cry1Ab9*, Lane 2: *cry1Ab14*, Lane 3: *cry 1Ea10*, Lane 4: *cry1Gc1*, Lane 5: *cry1Aa1a44*, Lane 6: *cry2Aa9*, Lane 7: *cry2Ab4*, Lane 8: *cry8Aa1*, lane9: *cry70Aa1*, Lane10: *cyt1Da2*, Lane 11: *vip3Aa59*, Lane 12: *vpb4Aa1*, Lane 13: *zwa5A*, Lane 14: *zwa5B*, Lane 15: *zwa6*

# Shatpada Armour (NBAIR Bt25)

## Scanning Electron Microscopy



## Multi Locus Sequence Typing

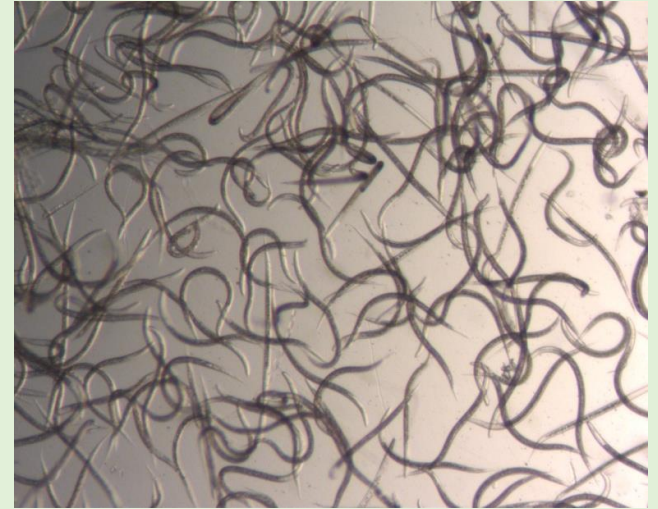


**Fig. 2:** Gel picture showing the amplification of housekeeping genes of *Bacillus thuringiensis* strain NBAIR BT25 in MLSA study. Lane M: Ladder (100bp). Lane 1: *glp*, Lane 2: *gmk*, Lane 3: *ilvD*, Lane 4: *ilvD2*, Lane 5: *pur*, Lane 6: *pta*, Lane 7: *pycA*, Lane 8: *tpi*



# Entomopathogenic nematodes

- *Heterorhabditis indica* 1 NBAIIH38 (ICAR NBAIR strain), *Steinernema carpocapsae* NBAIRS59 (ICAR NBAIR strain), ( $2.5 \times 10^8$  IJs ha<sup>-1</sup>) were tested.
- The second round of spraying of *H. indica* 1 NBAIIH38 (ICAR NBAIR Strain) at a concentration of  $2.5 \times 10^8$  IJs ha<sup>-1</sup>; Four releases of *T. chilonis* at weekly intervals (100 000 *T. chilonis* parasitized eggs ha<sup>-1</sup>) significantly reduced the larvae of *S. frugiperda* (87.56%), which is close to the value for emamectin benzoate treatment (98%).
- Technology approved for commercialization



# Trichogrammatid Cultures at ICAR-NBAIR

Live cultures (*Trichogrammatoidea* spp. (4 nos.) & *Trichogramma* spp. (20 nos.)

## *Trichogrammatoidea* spp.

*Trichogrammatoidea armigera* Manjunath

*Tr. sp. nr. robusta*

*Tr. bactrae* Nagaraja

*Tr. robusta* Nagaraja

## *Trichogramma* spp.

*Trichogramma achaeae* Nagaraja & Nagarkatti

*T. chilostraeae* Nagaraja & Nagarkatti

*T. hebbalensis* Nagaraja

*T. pieridis* Nagaraja & Mohanraj

*T. brassicae* Bezdenko (Italy)

*T. cacoeciae* Marchal (France)

*T. dendrolimi* Matsumura (Germany)

*T. evanescens* (Ar) (France)

*T. pretiosum* Riley (Colombia)

*T. pretiosum* (France)

*T. chilonis* Ishii

*T. danausida* Nagaraja

*T. japonicum* Ashmead

*T. semblidis* (Aurivillius) (France)

*T. brassicae* (Canada)

*T. cordubense* Vargas & Cabello (France)

*T. embryophagum* (Hartig) (Germany)

*T. evanescens* (Th) Westwood (France)

*T. mwanzai* Schulten & Feijen (Kenya)

*T. pretiosum* (Germany)

# Multiple insecticide tolerant strain of egg parasitoid *Trichogramma chilonis* for the management of FAW (NBAIR-MITS-FAW)

- The strain of the parasitoid, *Trichogramma chilonis* (MITS-FAW) is tolerant to multiple insecticides (organo-phosphate, pyrethroids, oxadiazine & spinosyn) with a high resistance factor and can be used against FAW and other pests in a pesticide stressed crop conditions including IPM.
- Technology approved for transfer.



Fall armyworm infesting  
maize



*T. chilonis* (MITS-FAW)  
parasitizing Fall armyworm eggs



# Awareness programmes

# GOI Initiatives.....

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कृषि, सहकारिता एवं किसान कल्याण विभाग  
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Government of India  
Ministry of Agriculture & Farmers Welfare  
Department of Agriculture, Cooperation  
& Farmers Welfare  
Krishi Bhawan, New Delhi-110001

Dated: 05.08.2019

*o/c*

**Subject: Management of Fall Armyworm in maize.**

We are aware that Fall Armyworm caterpillar a native to North America landed in Africa in 2016 and in India and many other Asian Countries in 2018. Since then, it has wreaked havoc in ravaging crops, especially maize. In fact, FAO has already declared FAW as a food security threat. In a short span, in India its presence has been observed in ten states and Fall Armyworm infestation has

**Management strategies of Fall Armyworm (FAW), *Spodoptera frugiperda* on maize**

**Monitoring:** Installation of pheromone traps @ 5/acre in the current and potential area of spread in crop season and off-season.

**Scouting:**  
Start scouting as soon as maize seedlings emerge

- At Seedling to early whorl stage (3-4 Weeks after emergence)- Action can be taken if 5% plants are damaged.
- At Mid whorl to late whorl stage (5-7 weeks after emergence)- Action can be taken if 10 % whorls are freshly damaged in mid whorl stage and 20% whorl damage in late whorl stage.
- At tassel and post-tasseling (Silking stage)- Do not spray chemical insecticides. Suitable bio-pesticide may be used in the event of ear/cob damage.

**Cultural Measures**

- Deep ploughing is recommended before sowing. This will expose FAW pupae to predators.
- Timely and uniform sowing over a large area is advised. Avoid staggered sowings.
- Intercropping of maize with suitable pulse crops of particular region. (eg. Maize + pigeon pea/black gram /green gram).
- Erection of bird perches @ 10/acre during early stage of the crop (up to 30 days).
- Sowing of 3-4 rows of trap crops (eg. Napier ) around maize field and spray with 5% NSKE or azadirachtin 1500 ppm as soon as the trap crop shows symptoms of FAW damage.
- Clean cultivation and balanced use of fertilizers.
- Cultivation of maize hybrids with tight husk cover will reduce ear damage by FAW.

**Mechanical control:**

- Hand picking and destruction of egg masses and neonate larvae in mass by crushing or immersing in kerosene water.
- Application of the seed in to the whorl of affected maize plants soon after observation of FAW incidence in the field.
- Application of Sand + lime in 9:1 ratio in whorls in first thirty days of sowing.
- Mass trapping of male moths using FAW specific pheromone traps @ 15/acre.

**Bio Control:**

- In crop protection of natural enemies by habitat management: Increase the plant diversity by intercropping with pulses, oil seeds and ornamental flowering plants which help in build-up of natural enemies.

• Regular surveys, surveillance and monitoring were conducted by the Central Integrated Pest Management Centres (CIPMCs) in collaboration with the State Department of Agriculture, SAUs and ICAR etc.

• Further, awareness programmes for the farmers were organized to advise them to adopt cultural and farm practices.

• Mass production of effective bio-control agents was promoted.

• The POP was circulated to all the States for its implementation. Timely advisories were issued to State Departments of Agriculture to adopt preventive measures.

# Efforts made by some State Agriculture Departments on management of FAW During 2022-23

State/UT	Details of Trainings held on FAW during 2022-23	
	No of Trainings Held	No of Farmers Participated
Andaman & Nicobar	12	350
Jharkhand	190	6850
Maharashtra	7245	118664
Mizoram	22	660
Punjab	231	7170
Uttar Pradesh	919	144675
Uttarakhand	24	4104



# Training activities conducted by CIPMCs of DPPQ&S, Govt of India

Year	Farmers Field School		2 days HRD		5 days HRD		30 days SLTP	
	Number	Farmers Trained	Number	# trained *	Number	# trained *	Number	Master Trainer trained
2019-20	690	21040	101	4120	11	440	3	120
2020-21	351	10530	28	1120	5	200	0	0
2021-22	282	9870	80	4368	0	0	0	0
2022-23 (Kharif)	126	4410	56	3063	0	0	0	0
2022-23 (Rabi)	104	3640	59	3174	6	229	0	0

\*AEOs, NGOs, Lead Farmers, Private Entrepreneurs etc.



# Distribution of NBAIR-BT25 formulations to farmers against FAW

**NBAIR-BT25 was distributed to farmers growing maize in Hindupur and Srikakulam for FAW management**





# Farmer Field days.....

- At Kadalaveni village, Gouribidanur Taluq, Chikkaballapur district, Karnataka.
- At Chilamattur, Hindupur, Karnataka.
- A large number of farmers field days were organized by state & central agencies.







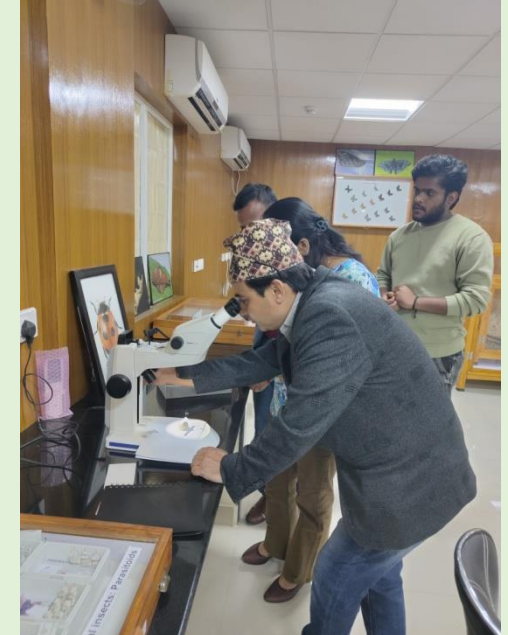
**Two days training on “Mass production of biocontrol agents for the management of fall armyworm (FAW)” (27-28 December 2019) under CABI funded project on FAW**





# International training programme on FAW

- Conducted **five days (22-26 May, 2023)** International training programme funded by **USAID** through the Feed the Future Nepal Integrated Pest Management (**FTFNIPM**) activity and the Feed the Future Bangladesh Integrated Pest Management (**IPMA**) activity being operated by, **Dr R Muniappan** Principal Investigator, FTFNIPM and IPMA, Virginia Tech, Blacksburg, USA.
- Seven officials from Nepal and Bangladesh were exposed to hands-on training on the “Production protocol of biocontrol agents for the management of Fall armyworm.”



# All India Coordinated research project on biological control of crop pests

## Validation of BIPM against FAW-Pan India

- Installation of pheromone trap @ 10 /acre
- Release of *Trichogramma chilonis* (1,00,000 eggs/ ha) (2 releases, first release after one week of sowing & second one after one week of first release )
- 2 sprays of NBAIR Bt-25 @ 10 ml/L at 10 days interval
- 2 sprays of NBAIR *Metarhizium anisopliae* (Ma-35) @ 5g/L at 10 days interval.
- C:B ratio: **1.77-4.78** at different locations





# Research Team at ICAR-NBAIR

*Thanks to the team members at ICAR-NBAIR for their dedicated service and sincere efforts.*



**Dr. A.N. Shylesha**



**Dr. A. Kandan**



**Dr. R. Rangeshwaran**



**Dr. Ankita Gupta**



**Dr. T. Venkatesan**



**Dr. Jagadeesh Patil**



**Dr. K Subaharan**



**Dr. Richa Varshney**



**Dr. G. Sivakumar**



**Dr. Omprakash Navik**



**Dr. Gandhi Gracy**

*Thank you  
for  
your kind attention*